



ELSEVIER

Contents lists available at ScienceDirect

Data in Brief

journal homepage: www.elsevier.com/locate/dib

Data Article

Data on the presence or absence of genes encoding essential proteins for ochratoxin and fumonisin biosynthesis in *Aspergillus niger* and *Aspergillus welwitschiae*

Fernanda Pelisson Massi ^{a,1}, Daniele Sartori ^{a,1},
 Larissa de Souza Ferranti ^{a,1}, Beatriz Thie Iamanaka ^{b,2},
 Marta Hiromi Taniwaki ^{b,2}, Maria Lucia Carneiro Vieira ^{c,3},
 Maria Helena Pelegrinelli Fungaro ^{a,*,1}

^a Centro de Ciências Biológicas, Universidade Estadual de Londrina, P.O. Box 6001, Londrina 86051-990, Brazil

^b Instituto de Tecnologia de Alimentos, P.O. Box 139, Campinas 13070-178, Brazil

^c Departamento de Genética, Escola Superior de Agricultura “Luiz de Queiroz” USP, P.O. Box 83, Piracicaba 13418-900, Brazil

ARTICLE INFO

Article history:

Received 18 January 2016

Received in revised form

18 February 2016

Accepted 3 March 2016

Available online 10 March 2016

Keywords:

Ochratoxin

Fumonisin

High-resolution capillary electrophoresis

ABSTRACT

We present the multiplex PCR data for the presence/absence of genes involved in OTA and FB₂ biosynthesis in *Aspergillus niger*/*Aspergillus welwitschiae* strains isolated from different food substrates in Brazil. Among the 175 strains analyzed, four mPCR profiles were found: Profile 1 (17%) highlights strains harboring in their genome the *pks*, *radH* and the *fum8* genes. Profile 2 (3.5%) highlights strains harboring genes involved in OTA biosynthesis i.e. *radH* and *pks*. Profile 3 (51.5%) highlights strains harboring the *fum8* gene. Profile 4 (28%) highlights strains not carrying the genes studied herein. This research content is supplemental to our original research article, “Prospecting for the incidence of genes

DOI of original article: <http://dx.doi.org/10.1016/j.ijfoodmicro.2016.01.010>

* Corresponding author.

E-mail address: fungaro@uel.br (M.H.P. Fungaro).

¹ Tel.: +55 43 3371 5731.

² Tel.: +55 19 3743 1819.

³ Tel.: +55 19 3429 4395.

<http://dx.doi.org/10.1016/j.dib.2016.03.016>

2352-3409/© 2016 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

involved in ochratoxin and fumonisin biosynthesis in Brazilian strains of *A. niger* and *A. welwitschiae*" [1].

© 2016 The Authors. Published by Elsevier Inc. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Specifications table

Subject area	Microbiology
More specific subject area	Mycology
Type of data	Figure
How data was acquired	Automated high-resolution capillary electrophoresis (CE). ABI 3500XL Genetic Analyser (Applied Biosystems, USA)
Data format	Analyzed
Experimental factors	A total of 175 strains of <i>Aspergillus niger</i> and <i>Aspergillus welwitschiae</i> isolated from different food substrates in distinct geographical regions of Brazil were submitted to DNA extraction and multiplex PCR analysis
Experimental features	Multiplex PCR using four primer pairs in one amplification reaction was carried out as described by Massi et al. [1]. The amplified products were analyzed using standard capillary electrophoresis
Data source location	Brazil
Data accessibility	The data is with this article

Value of the data

- The methodology presented here is potentially valuable to other researchers for developing similar assays for studying multiple genes simultaneously.
- Particularly, the multiplex PCR as presented here is useful to survey for the occurrence of *A. niger*/*A. welwitschiae* strains harboring essential genes for ochratoxin and fumonisin biosynthesis.
- The frequency of strains of *A. niger*/*A. welwitschiae* harboring essential genes for ochratoxin and fumonisin biosynthesis could be compared to that obtained from other countries.
- The multiplex PCR here developed is relevant to evidence specific non-producing mycotoxin phenotypes.

1. Data

Among 175 *A. niger*/*A. welwitschiae* strains analyzed, we found four mPCR profiles (Fig. 1). Profile 1 (17%) highlights strains harboring the *pks* (shown in blue, 554 bp), *radH* (blue, 328 bp) and *fum8* (blue, 128 bp) genes. Profile 2 (3.5%) highlights strains harboring only genes involved in OTA biosynthesis (*radH* and *pks*). Profile 3 (51.5%) highlights strains harboring only the gene *fum8*. Profile 4 (28%) highlights strains not carrying the mycotoxigenic genes studied herein.

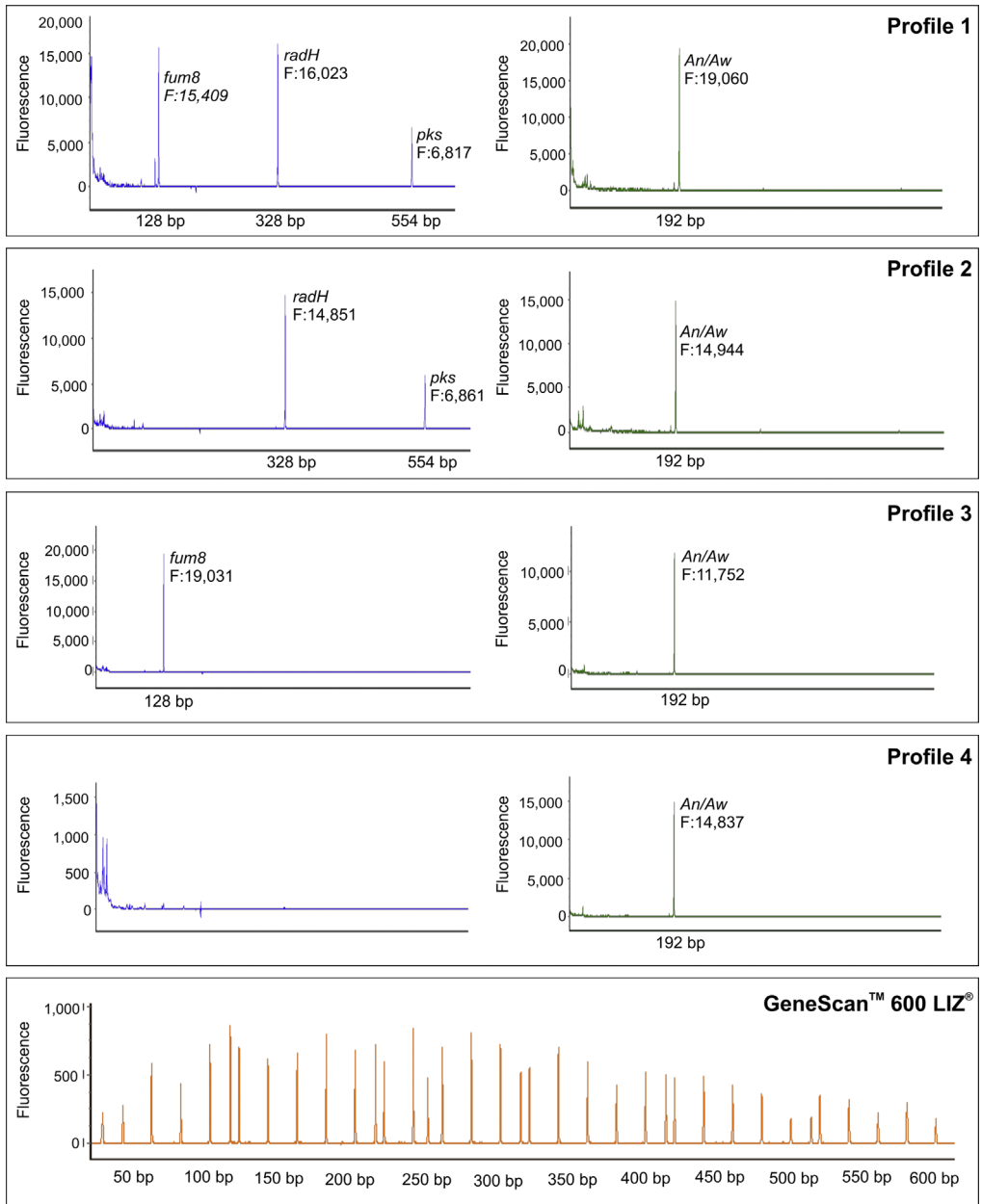


Fig. 1. Genotype profiles revealed by the ABI 3500XL Genetic Analyzer with GeneMarker® Software. Four PCR products were amplified with primers labeled with FAM (blue) and HEX (green). Profile 1 indicates that the strain is *A. niger* or *A. welwitschiae* (green, 192 bp) harboring the *pks* (blue, 554 bp), *radH* (blue, 328 bp) and the *fum8* (blue, 128 bp) genes. Profile 2 indicates that the strain is *A. niger* or *A. welwitschiae* harboring the genes involved in OTA biosynthesis i.e. *radH* and *pks*. Profile 3 indicates that the strain is *A. niger* or *A. welwitschiae* harboring the gene *fum8*. Profile 4 indicates that the strain is *A. niger* or *A. welwitschiae* harboring none of genes studied herein.



Fig. 2. Brazilian map depicting the locations from which samples were obtained.

2. Experimental design, materials and methods

We used a set of primer-pairs to survey for the presence/absence of genes involved in OTA and FB₂ biosynthesis in *A. niger*/*A. welwitschiae* strains [1], which were collected from dried fruits ($n=19$), Brazil nuts ($n=30$), coffee beans ($n=27$), grapes ($n=40$), cocoa ($n=3$), and onions ($n=56$). The Brazilian geographical regions where the samples were collected are shown in Fig. 2. The mycotoxigenic genes investigated herein were those encoding a polyketide synthase (*pks*), a flavin-dependent halogenase (*radH*), both involved in ochratoxin biosynthesis, and a α -oxoamine synthase (*fum8*), essential for fumonisin biosynthesis. A pair of *A. niger*/*A. welwitschiae*-specific primers targeting the β -tubulin gene (*benA*) was also included in the amplification reaction. Multiplex amplifications (*mPCR*) were carried out using four primer-pairs in a single reaction mixture, as described by Massi et al. [1]. Each amplified sample was diluted $10\times$ and 8.0 μL of (Hi-Di) formamide and 0.3 μL of GeneScan™ 600 LIZ[®] internal lane size standard (Applied Biosystems, USA) were added to 2 μL of the diluted sample. An ABI 3500XL Genetic Analyzer (Applied Biosystems, USA) was used to separate and detect the fluorescently labeled PCR products which were analyzed using GeneMarker[®] Software (SoftGenetics[®]).

Acknowledgments

This research was supported by the following Brazilian institutions: Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq 471813/2013-3), Fundação Araucária and Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP 2013/05414-8).

Appendix A. Supplementary material

Supplementary data associated with this article can be found in the online version at <http://dx.doi.org/10.1016/j.dib.2016.03.016>.

Reference

- [1] E.P. Massi, D. Sartori, L.S. Ferranti, B.T. Iamanaka, M.H. Taniwaki, M.L.C. Vieira, M.H.P. Fungaro, Prospecting for the incidence of genes involved in ochratoxin and fumonisin biosynthesis in Brazilian strains of *Aspergillus niger* and *Aspergillus wel-witschiae*, *Int. J. Food Microbiol.* 221 (2016) 19–28.